Phylogenetic Reconstruction: Handling Large Scale and Complex Data

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Acknowledgments

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Overview

- Phylogenies: What and Why?
- The Tree of Life and CIPRES
- Phylogenetic Reconstruction
- Scaling Up
- Gene Content and Order Data
  - Gene-Order Data: What and Why?
  - Computing with Gene-Order Data
  - Ancestral Gene Orders
- Summary
A phylogeny is a reconstruction of the evolutionary history of a collection of organisms.

It usually takes the form of a tree.

- Modern organisms are placed at the leaves.
- Edges denote evolutionary relationships.
- “Species” correspond to edge-disjoint paths.
The Great Apes

Phylogeny

From the Tree of the Life Website, University of Arizona

Orangutan  Gorilla  Chimpanzee  Human
Phylogenies: Why?

Phylogenies provide the framework around which to organize all biological and biomedical knowledge.

They help us understand and predict:

- functions of and interactions between genes
- relationship between genotype and phenotype
- host/parasite co-evolution
- drug targets
- origins and spread of disease
- origins and migrations of humans
Example: Antivenins

Venomous Australian Snakes

- Black whip snake
- Taipan
- Fierce snake
- Common brown
- Western brown
- Dugte
- Collatt's snake
- Spotted Black
- Butler's snake
- King brown
- Red-bellied black
- Death adder
- Barlick
- Small-eyed snake
- Australian copperhead
- Tiger snake
- Rough-scaled snake
- Broad-headed snake
Example: Antivenins

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Recommended Antivenin:
- Taipan
- Brownsnake
- Blcksnake
- Death adder
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The Tree of Life

It is to Biology what the periodic table is to Chemistry

BACTERIA

ARCHEA

EUKARYA

- Methanosarcina
- Methanobacterium
- Methanococcus
- Thermoplasma
- Thermococcus
- Chlamydiae
- Pyrodictium
- Thermoproteus
- Halophiles

- Thermotogales
- Thermophiles
- Flavobacteria
- Spirochetes
- Thermo cocci
- Deinococci
- Cyanobacteria
- Purple bacteria
- Gram-positive bacteria

- Diplomonads
- Ciliates
- Flagellates
- Fungi
- Entamoebae

- Ciliates
- Fungi
- Animals
- Plants
Scale of The Tree of Life

- 1,5 million described species.
- 10 million to 200 million existing species.
- Reconstruction tools can handle around 500 organisms.
- Reconstruction tools scale exponentially with the amount of data.
A community project funded for 5 years by the NSF for $12M under the ITR program, with the aim to develop the infrastructure (hardware, software, and databases) necessary to support the reconstruction of the Tree of Life.

- over 15 institutions, including three museums
- over 50 researchers, evenly split between CS and Biology
- research in algorithms, simulation and modelling, databases, software architecture, and high-performance computing
## CIPRES: Participants

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<tr>
<th>Institution</th>
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<tr>
<td>U. New Mexico</td>
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- Phylogenetic Reconstruction: *a fast review from a CS standpoint*
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Phylogenetic Reconstruction

Two categories of methods:

- **Criterion-Based** methods, such as Maximum Parsimony (MP) and Maximum Likelihood (ML)
- **Ad hoc**, usually *distance-based* and using clustering ideas, such as Neighbor-Joining (NJ)

**In addition:**

- **Meta-methods** decompose the data into smaller subsets, construct trees on those subsets, and use the resulting trees to build a tree for the entire dataset (quartets, disk-covering)
Phylogenetic Distances

- **True evolutionary distance:**
  the *actual* number of evolutionary events that took place to transform one datum into the other.

- **Edit distance:**
  the *minimum* number of permitted evolutionary events that can transform one datum into the other.

- **Estimated evolutionary distance:**
  our best *estimate* of the true evolutionary distance, obtained heuristically or by correcting the edit distance according to a model of evolution.
Distance-Based Methods

*(Unknown) True Tree*

- A
- B
- C
- D
- E

Extract data on extant taxa

Inferred Tree

- A
- B
- C
- D
- E

Distance Matrix

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Molecular Data

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Aim to minimize total *number of character changes*. Assume that characters are *independent*. Reconstruct *ancestral data*. Finding a most parsimonious tree is NP-hard.

Optimal solutions are limited to sizes around 30. Heuristic solutions appear good to about 500 taxa (e.g., TNT).
Likelihood-Based Methods

Aim to return tree with highest likelihood of having produced the observed data.

Are based on a specific model of evolution and usually estimate model parameters.

Produce likelihood estimate (prior or posterior conditional) for each tree.

Even scoring a fixed tree is very expensive.

Optimal solutions are limited to specific sets of 4 taxa. Heuristics run to completion on at most 15 taxa, but appear good to about 100 taxa (e.g., MrBayes, PhyML, RAxML).
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Scaling Up: The Issues

- *Distance-based methods are (fairly) fast, but not accurate enough on large problems (large evolutionary diameter).*
- *Criterion-based methods take days for a few hundred taxa and scale exponentially.*
- *All methods perform better with longer sequences and larger state spaces, but biological sequences are bounded.*
Scaling Up: Approaches

- *Distance-based methods are (fairly) fast, but not accurate enough on large problems.*
  Decompose large problems into smaller ones so as to reduce evolutionary diameter.

- *Criterion-based methods take days for a few hundred taxa and scale exponentially.*
  Use algorithmic techniques to bypass the exponential growth, such as divide-and-conquer.

- *All methods perform better with longer sequences and larger state spaces, but biological sequences are bounded.*
  Design methods that converge on short sequences, so-called *fast converging methods.*
Scaling Up: A Solution

Disk-Covering Methods

Basic idea:

- decompose dataset into *overlapping* compact subsets—the *disks*
- reconstruct a tree for each subset
- assemble these trees into a single tree

Variations so far: DCM1, DCM2, DCM3, recursive versions, iterative versions
DCM1 and DCM2

DCM1

4 disks

DCM2

separator in green
Improvement: DCM3

DCM1 and DCM2: decomposition based on distance matrix only

DCM3: use best tree so far to guide the decomposition

Given set $S$ and tree $T$, compute short subtree graph $G(S,T)$ and find clique separator in $G$ to form subproblems.
Using DCM3: Recurse & Iterate

1. Obtain initial guide tree
2. If satisfied with tree?
   a. Output tree
   b. Refine tree
     i. Assemble subtrees into full tree
     ii. Solve final subproblems with chosen base method
   c. Replace guide tree by new tree
     i. Using guide tree, recursively decompose down to desired size
Results with DCM1 and NJ

using Kimura 2-parameter plus $\Gamma$ model

reduced sequence length
(0.15 error rate)

reduced error rate
(1,000 sequence length)
Results with Rec-I-DCM3 and MP

Rec-I-DCM3(TNT) vs. TNT

10,000 RNA sequences

10 datasets
(from 4,000 to 15,000)

Finding: 0.01% error is the maximum allowed!!
Results with Rec-I-DCM3 and MP

**Rec-I-DCM3(TNT) vs. TNT**

10,000 RNA sequences

Finding: 0.01% error is the maximum allowed!!
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Phylogenetic Data

- All kinds of data have been used: behavioral, morphological, metabolic, etc.
- Current data of choice are molecular data.
- Two main kinds of molecular data:
  - **sequence**
    (nucleotide/codon sequences from genes)
  - **gene content and order**
    (gene ordering on chromosomes)
Gene-Order Data

The ordered sequence of genes on one or more chromosomes.

The entire gene order is a *single character*, which can assume a huge number of states.

Evolves through *inversions*, *insertions* (incl. duplications), and *deletions*; also *transpositions* (seen in mitochondria) and *translocations* (between chromosomes).

- Need to identify genes and gene families.
- Need to refine model for specific organisms to handle operons, exons, etc.
Genome Rearrangements

Model based on three types of rearrangements:

- **Inversion**
- **Transposition**
- **Inverted Transposition**

![Diagram of genome rearrangements with numbers and arrows indicating the transformations.](image-url)
Gene-Order Data: Attributes

Advantages:

- Rare genomic events (*sensu* Rokas/Holland) and huge state space, so very low risk of homoplasy.
- No need for alignments.
- No gene tree/species tree problem.

Problems:

- Mathematics *much more complex* than for sequence data.
- Models of evolution not well characterized.
- Very limited data (mostly organelles and bacteria).
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Breakpoint Distance

The number of adjacencies present in one genome, but not the other.

\[ G_1 = (1 \ 2 \ 3 \ 4 \ 5 \ 6 \ 7 \ 8) \]

\[ G_2 = (1 \ 2 \ -5 \ -4 \ -3 \ 6 \ 7 \ 8) \]
Gene-Order Distances in General

Signed gene orders may include duplicates, need not have identical gene content.

Previous work (not useable for phylogeny):

- Exemplar heuristic for duplications by Sankoff (NP-hard).
- Exact inversions plus deletions, but no duplications allowed, by El-Mabrouk.
- Heuristic by Bourque, used only on very small sets.

Our work:

- Bounded approximation for unequal gene content.
- Direct estimate of evolutionary distance.
Direct Distance Estimate: Example

Simulated 800-gene genomes, 70% inversions (mean of 20, located uniformly), 16% deletions, 7% insertions, and 7% duplications (all mean 10).

*left: expected pairwise distances from 40 to 160 events*
*right: expected pairwise distances from 80 to 320 events*
Using Our Distance Estimate

(unpublished)

13 gamma proteobacteria (Lerat/Daubin/Moran 2003)
Over 3,400 genes, with 540–3,000 genes and 3%–30% duplications per genome; pairwise distances from 170 to 1700 events.

Reference phylogeny: 2 years of work, over 60 gene sequences.

Using our distance estimates and naïve NJ:
1 hour to compute distances, 1 second to construct tree, and only one error (long branch attraction, trivially fixed).
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**Goal:** Reconstruct a signed gene order at each internal node in the tree to minimize sum of edge distances.

Problem is NP-hard even for just three leaves, no duplications, and simplest of distances (breakpoint, plain inversion)!

This is the **median problem** for signed genomes: given three genomes, produce a new genome that will minimize the sum of the distances from it to the other three.
Observations on Ancestral Genomes

- Successful for small, streamlined genomes (organelles).
- Must take into account gene content: common simplification of reducing to just shared genes loses too much information.
- Not doable for bacterial nuclear genomes without additional biological constraints: too many solutions.
- Possible additional constraints: hot spots, lengths of affected segments, protected segments (centromere, origin of replication, etc.), nucleotide data around each gene, etc.
Medians with Unequal Gene Content

Tang/Moret/Cui/DePamphilis (2004): chloroplast data

organismal

NJ (inv.)

Tang/Moret GRAPPA

breakpoint GRAPPA
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- Complex Data: Gene-order data carry a strong phylogenetic signal and current algorithmic approaches scale to significant sizes.
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- Complex Data: Gene-order data carry a strong phylogenetic signal and current algorithmic approaches scale to significant sizes.

- Approach: Strong algorithmic design, constant iteration on evolutionary models, extensive testing on simulated data and biological data. Stimulate CS research (even if highly abstract) and biological research.
Thank You!

Laboratory for High-Performance Algorithm Engineering and Computational Molecular Biology

compbio.unm.edu

CIPRES
Cyber Infrastructure for Phylogenetic Research

www.phylo.org